

Contents:	Cell counter (Sysmex K-1000) values. Cell counting was performed with a Sysmex K-1000, with related computer part DA 1000, Toa Medical Electronics, Japan, purchased in 1991. Data were generated from four main qualifiers (type of sample), all individuals have results from whole blood (QUAL=0). A marker (minus sign, "-") in front of all abnormal values (outside the reference range) was removed in 2023.
# lines:	76 305 (30 362 individuals)
# variables:	18
Reference:	<p>Adamsson Eryd S, Smith J G, Melander O, Hedblad B, Engström G. Incidence of coronary events and case fatality rate in relation to blood lymphocyte and neutrophil counts. <i>Arterioscler Thromb Vasc Biol.</i> 2012;32:533-539</p> <p>Borné Y, Smith J G, Melander O, Hedblad B, Engström G. Red cell distribution width and risk for first hospitalization due to heart failure: a population-based cohort study. <i>Eur J Heart Failure.</i> 2011;13:1355-1361.</p> <p>Martinsson A, Andersson C, Andell P, Koul S, Engstrom G, Smith J G. Anemia in the general population: prevalence, clinical correlates and prognostic impact. <i>Eur J Epidemiol.</i> 2014;29:489–498.</p> <p>Patti P, Di Martino G, Ricci F, Renda G, Gallina S, Hamrefors V, Melander O, Sutton R, Engström G, De Caterina R, Fedorowski A. Platelet Indices and Risk of Death and Cardiovascular Events: Results from a Large Population-Based Cohort Study. <i>Thromb Haemost.</i> 2019 Nov;119(11):1773-1784. doi: 10.1055/s-0039-1694969.</p> <p>Sysmex operators manual k-1000. Kobe, Japan. <i>Toa Medical Electronics Co. Ltd.</i> 1993. 150 s.</p> <p>Sysmex k-1000 specifications. Kobe, Japan. <i>Toa Medical Electronics Co. Ltd.</i> 1 s.</p>

List of variables

Name	Variable label	Type	Format	Value label
lopnrMKC	Baseline sequence number in MDCS (Numeric)	Numeric	F5	
labnrMKC	Baseline sequence number of laboratory sample in MDCS (originally named LABNO)	Numeric	F5	
QUAL	Qualifiers, type of sample (QUAL=0 is usually used)	Numeric	F1	0 = Whole blood, (differential count on heparin blood), available from all participants 1 = Isolated mononuclear leukocytes, available from 16000 participants 2 = Isolated granulocytes (GRAN), available from the first 16000 participants 3 = Erythrocytes (quality control) 4 = EDTA blood (about 200 participants, analysed by Sölve/ S Anders) 5 6 = Buffy coat (WBC, leukocytes), available from the last 14000 participants 7 8 9
DATUM	Date at sample collection at MDC baseline	Numeric	F6	
WBC_23	Leukocyte count, white blood cells (x10 ⁹ cells/liter). <i>Remarks:</i> WBC= LYMPH + MXD + NEUT , where denotes the absolute value. The relationship is valid for all records (with few exceptions) with values for LYMPH, MXD and NEUT.	Numeric	F5.2	
LYMPH_23	Lymphocyte count (x10 ⁹ cells/liter)	Numeric	F5.2	

Name	Variable label	Type	Format	Value label
MXD_23	Mixed cell (monocyte rich) count (x10 ⁹ cells/liter)	Numeric	F5.2	
NEUT_23	Neutrophil (granulocyte) count (x10 ⁹ cells/liter)	Numeric	F5.1	
RBC_23	Erythrocyte count, red blood cells (x10 ¹² cells/liter)	Numeric	F4.2	
SD_23	Erythrocyte distribution width	Numeric	F5.1	
HGB_23	Hemoglobin concentration (g/liter)	Numeric	F3	
HCT_23	Hematocrit, true relative percentage volume of erythrocytes (%)	Numeric	F5.2	
MCV_23	Mean erythrocyte volume (fl, femto liter, 10 ⁻¹⁵ liter)	Numeric	F5.1	
MCH_23	Mean erythrocyte hemoglobin (pg, pico gram, 10 ⁻¹² gram)	Numeric	F5.1	
MCHC_23	Mean erythrocyte hemoglobin concentration (g/liter)	Numeric	F4	
PLT_23	Platelet count (number of particles * 10 ⁹ /liter)	Numeric	F4	
PDW_23	Platelet distribution width (fl, femto liter, 10 ⁻¹⁵ liter)	Numeric	F5.2	
MPW_23	Mean platelet volume (fl, femto liter, 10 ⁻¹⁵ liter)	Numeric	F5.2	